Hypoxia-targeted bioreductive tyrosine kinase inhibitors with glutathione-depleting function

Rangaprasad Sarangarajan^a, Shireesh P. Apte^b and Sydney O. Ugwu^c

Tyrosine kinase inhibitors may serve as ligands for kinases that are involved in normal cell differentiation or repair, thereby leading to toxicity. It may be possible to target such inhibitors to tumor cells by coupling them to hypoxia-activated bioreductive molecules. Such coupling can utilize or incorporate bonds that have a propensity to be preferentially oxidized by thiols such as intracellular glutathione (GSH). The resulting depletion of GSH may increase redox-mediated apoptosis. The resultant molecule is hence projected to act via multiple cell killing mechanisms: (i) inhibition of tumor kinases, (ii) tumor DNA disruption and (iii) causing increased redox-mediated apoptosis. Anti-Cancer Drugs 17:21-24 © 2006 Lippincott Williams & Wilkins.

The epidermal growth factor receptor (EGFR) is a molecular target of cancer therapies because it is overexpressed in various epithelial tumors [1]. An EGFR inhibitor used as a monotherapy in non-small cell lung cancer (NSCLC) however, has been associated with interstitial lung fibrosis. The inappropriate regeneration of sequentially injured epithelium represents an important process leading to pulmonary fibrosis [2]. Previous studies have demonstrated that epithelial expression of EGFR is upregulated in fibrotic lung tissue, suggesting that EGFR-mediated signaling is involved in epithelial regeneration in fibrotic lung disease. Thus, in NSCLC patients with underlying or a predisposition to lung injury, inhibition of the EGFR may also trigger interstitial pulmonary fibrosis. In addition, EFGR inhibition may also augment fibrosis by stimulating apoptosis in alveolar epithelial cells [3,4], by inhibiting epithelial differentiation [5] or by inhibiting pulmonary angiogenesis [6].

Members of the vascular endothelial growth factor (VEGF) play a critical role in angiogenesis [7]. Therefore, the disruption of the VEGF pathway represents an attractive target for anti-cancer therapy. Angiogenesis inhibitors that target VEGF have been associated with increased incidences of thromboembolic events [8]. VEGF has been shown to be involved in hemostasis by altering the hemostatic properties of endothelial cells [9]. A low basal level of VEGF activity has been postulated to keep nontumoral endothelial cells in a non-activated state, partly by inducing nitric oxide synthetase expression and nitric oxide production [10,11]. Blockage of endothelial VEGF results in significant elevations of s-E-selectin and s-tissue factor reflecting a pro-thrombotic state and a decrease in the level of the coagulation cascade. The pro-thrombotic

Anti-Cancer Drugs 2006, 17:21-24

Keywords: bioreductive, hypoxia, inhibitor, glutathione, thiol, tyrosine kinase

^aDepartment of Pharmaceutical Sciences, Massachusetts College of Pharmacy and Health Sciences, Worcester, Massachusetts, ^bBaxter I.V. Systems, Murray Hill, New Jersey and ^cNeopharm Inc, Waukegan, Illinois, USA.

Correspondence to S.P. Apte, 2313 Welch Place, Mansfield, TX 76063, USA. Tel: +1 817 689 6086 or 817 453 1898; fax +1 817 551-8626; e-mail: shireeshpapte@msn.com

Received 3 May 2005 Accepted 30 August 2005

state is aggravated by concomitant administration of cytotoxic agents that induce thrombocytopenia because platelets are carriers of VEGF [12].

It is now becoming evident that therapies that target multiple targets or signaling pathways in malignant diseases are likely to be more efficacious than receptoror molecule-specific drugs [13-15]. In particular, the ability of kinases to mutate in response to the selective pressure created by treatment with tyrosine kinase inhibitors (TKIs) alone provides a strong rationale of hitting more than one essential target at the same time in the tumor cells [16]. This can be achieved, for example, by linking TKIs to hypoxia-activated pro-drugs or by engineering a single TKI to inhibit multiple target receptors. For example, SU6668 is a potent inhibitor of VEGF receptor-2, platelet-derived growth factor receptor (PDGFR) and fibroblast growth factor receptor [17] three transmembrane tyrosine kinases involved in different phases of tumor angiogenesis. Imatinib mesylate (Gleevec) targets the Abl tyrosine kinase, c-kit (stem cell factor receptor) as well as PDGFR tyrosine kinases [18]. The better clinical outcomes and relative non-toxic profile of these drugs may be attributed in part to their molecular 'promiscuity' against multiple targets and blocking multiple steps in tumor growth. As more ATP competitive inhibitors are developed, however, it is possible that some of these may also serve as ligands for other kinases whose inactivation may result in undue host toxicity.

How can such drugs be made to 'differentiate' between cells that are proliferating due to impaired apoptotic

0959-4973 © 2006 Lippincott Williams & Wilkins

control mechanisms (tumors) versus those that are dividing (i) either normally or (ii) as a response to cellular or organ injury or microbial attack? For example, such a TKI must be able to differentiate between overexpressed EGFR in tumors versus overexpressed EGFR that serves to ameliorate pulmonary fibrosis and low basal levels of EGFR signaling by normally functioning quiescent endothelial cells. Such 'differentiation' between tumor cells and normally dividing cells may be achieved by mechanisms that serve to target the TKI to tumor cells alone. One approach is to link the TKI to a hypoxia-activated pro-drug.

In 1924, Otto Warberg hypothesized that 'the prime cause of cancer is the replacement of the respiration of oxygen in normal body cells by a fermentation of sugar' [19]. Aerobic glycolysis has since been shown to be elevated in over 90% of all metastatic cancers [20]. The products of such glycolysis, lactate and pyruvate, have been linked to a lower extracellular tumor pH and to a stimulation of the accumulation of hypoxia-inducible factor (HIF)- 1α [21]. The resultant hypoxia induces proteomic and genomic changes within the tumor cells that result in malignant progression and treatment resistance [22,23].

Low oxygen tensions distinguish hypoxic tumor cells from normal cells [24]. Therefore, it may reasonably be assumed that the redox environment of tumors is different from that of normal cells. Indeed, it has been demonstrated that an electron-rich reducing environment exists in tumors [25] along with an elevated concentration of the primary intracellular buffer – glutathione (GSH) [26]. GSH has been implicated in protection against the induction of apoptotic and necrotic cell death in a variety of cell types [27–29], in part by redox modulation of cytochrome c and caspase release [30,31]. Depletion of cellular reduced GSH has also been shown to abolish the multidrug resistance-associated protein MRP1-mediated resistance against vinca alkaloids and anthracyclines, thereby making GSH-depleted cells more vulnerable to chemotherapy [32,33]. It has been speculated that reducing the elevated levels of GSH in tumors could inhibit aerobic glycolysis, which in turn may result in normalization of oxygen perfusion and pH [34]. Consequently, the depletion of GSH in hypoxic tumor tissue may have beneficial effects on the treatment of tumors [35,36]. GSH-influenced intracellular redox state is coupled to the oxidation state of cysteine residues in proteins [37] by complex thiol/disulfide exchange mechanisms that in turn influence the activity of a variety of enzymes [38].

Hypoxia-selective bioreductive pro-drugs, rather than undergoing selective reduction, undergo reduction events in all cells. These reductive events are reversible in the presence of molecular oxygen (via superoxide generation) [37], but irreversible in its absence. Excellent reviews of hypoxia-activated drug design can be found [39-44]. In hypoxic tumors, such pro-drugs are reduced by ubiquitous cellular reductases (cytochrome P450 reductase, xanthine oxidase) to yield electrophilic traps for DNA amino acid nucleophiles [44]. Since the efficiency of redox cycling depends upon the rate of catalytic turnover, optimum activity results when the one-electron redox potential of the pro-drug is in between that of cellular reductants, around -0.4V versus normal hydrogen electrode (NHE) and that of the $O_2/O_2^{-\bullet}$, around -0.15 V versus NHE in an aqueous buffered medium, at pH 7.0 [45]. Molecules with too low a rate of reduction (low electron affinities) are expected to be inadequately activated by bioreductive enzymes even under hypoxic conditions, while molecules with too high a rate of reduction (high electron affinity) when compared with the rate of radical reactivity with oxygen will be cytotoxic toward normo-oxic tissue. It would hence appear that molecules with electron affinities less than -0.35 and greater than -0.15 V will show little activity against hypoxic cells in vivo.

The concept that GSH (as opposed to cellular reductase enzymes) can be used to reduce hypoxia-selective bioreductive pro-drugs has not received much attention.

The redox potential of the GSSG/2GSH couple is -0.25 V. Therefore, a bioreductive pro-drug may be preferentially reduced by cellular reductases rather than by GSH due to the larger redox potential difference between the pro-drug and the reductase than between the pro-drug and GSH. Furthermore, a bioreductive pro-drug whose redox potential falls between -0.25(GSSG/2GSH) and -0.15 V ($O_2/O_2^{-\bullet}$) could potentially be reduced (and consequently be cytotoxic) in normooxic tissue. Even if a pro-drug could be designed so that its redox potential is lower at physiological pH that exists in normo-oxic tissue, but increases at the lower pH that exists in hypoxic tissue, it would still be preferentially reduced by cellular reductases instead of by GSH. Therefore, modulation of redox potential for design of a hypoxia-specific GSH reducible cytotoxin does not seem particularly attractive.

Another approach would be to design pro-drugs that are reducible preferentially by thiols. For example, KW-2149 (an analog of mitomycin C) is activated by non-protein thiols rather than by bioreductive enzymes, presumable due to the presence of a disulfide bond [46]. A novel class of Pt(IV) dicarboxylate compounds has been developed which are preferentially reduced and activated by thiols [47] occurring via a reductive elimination process through an attack by sulfur at one of the mutually trans chloride ligands [48]. A heteroaromatic N-oxide, tirapazamine, is the first hypoxia-selective pro-drug registered for clinical use [49]. Its low aqueous solubility however, and slow extracellular diffusion have prompted a search for analogs

with more favorable physicochemical characteristics [50]. Linking this molecule to a suitable hydrophilic TKI by a disulfide bond as well as imparting a more acidic character may increase opportunities for favorable modulation of the physicochemical properties as well as inducing thiol-specific reductivity. Some possible constructs are shown in Fig. 1. Figure 1(I) shows such a possible tirapazamine-TKI construct, linked by a disulfide bond. Figure 1(II) shows the possibility of modulating the solubility and diffusion kinetics of Fig. 1(I) by attachment of a platinum pro-drug (JM335) [48] to the N-oxide ring of tirapazamine. The platinum chelate was chosen because it can also (in addition to the disulfide bond) be reduced by thiols, thereby increasing the molecule's GSH-depleting ability.

It should be emphasized that the above constructs represent 'thought experiments'. The authors have no data to show that such molecules will actually be effective as envisaged in vivo. Nevertheless, these structures may serve as good first approximations for the design and testing of the hypoxia-mediated TKI paradigm. It is also evident that cleavage of the disulfide bond by GSH will admittedly leave a residual thiol group on the released TKI molecule. However, if positioned away from the catalytic ATP-binding site, such an added -SH group may not be detrimental to the TKI-binding affinity.

Such a redox cytotoxic entity linked through a disulfide bond that is cleavable by thiols such as GSH to a TKI would (i) selectively target the TKI to tumors, thereby improving safety, decreasing TKI drug resistance and increasing the therapeutic index, (ii) cause a depletion of tumor GSH levels causing redox-mediated apoptosis and/ or increased susceptibility to reactive oxygen species (ROS), and (iii) release ROS in the vicinity of tumor cells causing DNA damage.

It has been shown that it is possible to release phenols, amines, thiols and carboxylic acids from appropriate bioreductive indolequinones selectively in hypoxic tissue [51]. Proof-of-concept has been obtained with pro-drugs comprising a bioreducible 'trigger' linked to an 'effector', which fragments off from the intermediate radical [52]. It is therefore reasonable to assume that the release of small-molecule protein kinase inhibitors could be selectively accomplished in hypoxic tumor tissue by linkage with suitable bioreductive agents.

Conclusions

TKIs may be targeted to tumors by linkage with hypoxiaactivated pro-drugs. The linker can be a bond that is preferentially degraded by intracellular thiols such as GSH. The redox and physicochemical properties of the hypoxia-activated pro-drug may be more amenable to manipulation in the presence of the TKI, leading to more diffusion into tumor tissue and increased selective hypoxia cytotoxicity. Conversely, the hypoxia-activated pro-drug serves to target the TKI and the linker serves to deplete intracellular GSH, thereby possibly rendering

Fig. 1

$$\begin{array}{c} O \\ \downarrow \\ N^{+} \\ \downarrow \\ \\ N^{$$

Tyrosine kinase inhibitors linked to hypoxia activated bioreductive prodrugs via glutathione depleting disulfide bonds.

the tumor tissue more susceptible to redox-mediated apoptosis and decreasing TKI resistance.

References

- Ciardiello F, Tortora G. A novel approach in the treatment of cancer: targeting the epidermal growth factor receptor. Clin Cancer Res 2001;
- Selman M, King Jr TE, Pardo A. Idiopathic pulmonary fibrosis: prevailing and evolving hypotheses about its pathogenesis and implications for therapy. Ann Intern Med 2001: 134:136-151.
- 3 Ciardiello F, Caputo R, Bianco R, Damiano V, Pomatico G, De Placido S, et al. Antitumor effect and potentiation of cytotoxic drug activity in human cancer cells by ZD1839 (Iressa), an epidermal growth factor receptorselective tyrosine kinase inhibitor. Clin Cancer Res 2000; 6:2053-2063.
- Kuwano K, Hagimoto N, Kawasaki M, Yatomi T, Nakamura N, Nagata S, et al. Essential roles of the Fas-Fas ligand pathway in the development of pulmonary fibrosis. J Clin Invest 1999; 104:13-19.
- Yasui S, Nagai A, Oohira A, Iwashita M, Konno K, et al. Effects of anti-mouse EGF antiserum on prenatal lung development in fetal mice. Pediatr Pulmonol 1993; 15:251-256.
- Ciardiello F, Caputo R, Bianco R, Damiano V, Fontanini G, Cuccato S, et al. Inhibition of growth factor production and angiogenesis in human cancer cells by ZD1839 (Iressa), a selective epidermal growth factor receptor tyrosine kinase inhibitor. Clin Cancer Res 2001; 7:1459-1465.
- Carmeliet P, Jain RK. Angiogenesis in cancer and other diseases. Nature 2000; 407:249-257.
- Daly ME, Makris A, Reed M, Lewis CE. Hemostatic regulators of tumor angiogenesis: a source of antiangiogenic agents for cancer treatment? J Natl Cancer Inst 2003; 95:1660-1673.
- Kuenen BC, Levi M, Meijers JCM, Kakkar AK, van Hinsbergh VWM, Kostense PJ. Analysis of coagulation cascade and endothelial cell activation during inhibition of vascular endothelial growth factor/vascular endothelial growth factor receptor pathway in cancer patients. Arterioscler Thromb Vasc Biol 2002: 22:1500-1505.
- 10 Shen BQ, Lee DY, Zioncheck TF. Vascular endothelial growth factor governs endothelial nitric oxide synthase expression via a KDR/ Flk-1 receptor and a protein kinase C signaling pathway. J Biol Chem 1999; 274:33057-33063.
- Van der Zee R, Murohara T, Luo Z, Zollmann F, Passeri J, Lekutat C, et al. Vascular endothelial growth factor/vascular permeability factor augments nitric oxide release from quiescent rabbit and human vascular endothelium. Circulation 1997; 95:1030-1037.
- Kuenen BC, Levi M, Meijers JCM, van Hinsbergh VWM, Berkhof J, Kakkar AK, et al. Potential role of platelets in endothelial damage observed during treatment with cisplatin, gemcitabine, and the angiogenesis inhibitor SU5416. J Clin Oncol 2003: 21:2192-2198.
- O'Reilly M. Targeting multiple biological pathways as a strategy to improve the treatment of cancer. Clin Cancer Res 2002; 8:3309-3310.
- Arteaga CL. Molecular therapeutics: is one promiscuous drug against multiple targets better then combinations of molecule-specific drugs? Clin Cancer Res 2003; 9:1231-1232.
- 15 DuBois RN. New paradigms for cancer prevention. Carcinogenesis 2001; 22:691-692
- Druker BJ. Overcoming resistance to imatinib by combining targeted agents. Mol Cancer Ther 2003; 2:225-226.
- Laird AD, Vajkoczy P, Shawver LK, Thurnher A, Liang C, Mohammadi M, et al. SU6668 is a potent antiangiogenic and antitumor agent that induces regression of established tumors. Cancer Res 2000: 60:4152-4160.
- Buchdunger E, Cioffi CL, Law N, Stover D, Ohno-Jones S, Druker BJ, et al. Abl protein-tyrosine kinase inhibitor STI571 inhibits in vitro signal transduction mediated by c-kit and platelet derived growth factor receptors. J Pharmacol Exp Ther 2000; 295:139-145.
- 19 Warburg O. The metabolism of tumors. London: Arnold Constable; 1930.
- Gambhir SS, Czernin J, Schwimmer J, Silverman DH, Coleman RE, Phelps ME. A tabulated summary of the FDG PET literature. J Nucl Med 2001; 42:1S-93S.
- 21 Lu H, Forbes RA, Verma A. Hypoxia-inducible factor 1 activation by aerobic glycolysis implicates the Warburg effect in carcinogenesis. J Biol Chem 2002; 277:23111-23115.
- Vaupel P, Thews O, Hoeckel M. Treatment resistance of solid tumors: role of hypoxia and anemia. Med Oncol 2001; 18:243-259.
- Hockel M, Vaupel P. Tumor hypoxia: definitions and current clinical, biologic and molecular aspects. J Natl Cancer Inst 2001; 93:266-276.
- Vaupel P, Harrison L. Tumor hypoxia: causative factors, compensatory mechanisms, and cellular response. The Oncologist 2004; 9:4-9.

- 25 Ilangovan G, Li H, Zweier JL, Kuppusamy P. In vivo measurement of tumor redox environment using EPR spectroscopy. Mol Cell Biochem 2002; 234-235:393-398.
- 26 Kuppusamy P, Li H, Ilangovan G, Cardounel AJ, Zweier JL, Yamada K, et al. Noninvasive imaging of tumor redox status and its modification by tissue glutathione levels. Cancer Res 2002; 62:307-312.
- Hockenbery DM, Oltvai ZN, Yin X, Milliman CL, Korsmeyer SJ. Bcl-2 functions in an antioxidant pathway to prevent apoptosis. Cell 1993; 75:241-251.
- Kane DJ, Sarafian TA, Anton R, Hahn H, Gralla EB, Valentine JS, et al. Bcl-2 inhibition of neural death; decreased generation of reactive oxygen species. Science 1993; 262:1274-1277.
- Adamson GM, Billings RE. Tumor necrosis factor induced oxidative stress in isolated mouse hepatocytes. Arch Biochem Biophys 1992;
- Ghibelli L, Coppola S, Fanelli C, Rotilio G, Civitareale P, Scovassi Al, et al. Glutathione depletion causes cytochrome c release even in the absence of cell commitment to apoptosis. FASEB J 1999; 13:2031-2036.
- Kirkland RA, Franklin JL. Evidence for redox regulation of cytochrome c release during programmed neuronal death: antioxidant effects of protein synthesis and caspase inhibition. J Neurosci 2001; 21:1949-1963.
- Zaman GJ, Lankelma J, van Tellingen O, Beijnen J, Dekker H, Paulusma C, et al. Role of glutathione in the export of compounds from cells by the multidrug resistance associated protein. Proc Natl Acad Sci USA 1995; 92:7690-7694.
- Versantvoort CH, Broxterman HJ, Bagrij T, Scheper RJ, Twentyman PR. Regulation by glutathione of drug transport in multidrug-resistant human lung tumor cell lines overexpressing multidrug resistance-associated protein. Br J Cancer 1995; 72:82-89.
- Benabe JE, Echegoyen LA, Pastrana B, Martinez-Maldonado M. Mechanism of inhibition of glycolysis by vanadate. J Biol Chem 1987; 262:9555-9560.
- Meister A. Glutathione deficiency produced by inhibition of its synthesis, and its reversal; applications in research and therapy. Pharmacol Ther 1991;
- O'Dwyer, Hamilton PJ, LaCreta TC, Gallo JM, Kilpatrick D, Halbherr T, et al. Phase I trial of buthionine sulfoximine in combination with melphalan in patients with cancer. I Clin Oncol 1996: 14:249-256.
- Ziegler DM. Role of reversible oxidation-reduction of enzyme thiols-disulfides in metabolic regulation. Annu Rev Biochem 1985; 54:305-329.
- Gilbert HF. Redox control of enzyme activities by thiol/disulfide exchange. Methods Enzymol 1984; 107:330-351.
- Stratford IJ, Workman P. Bioreductive drugs into the next millennium. Anticancer Drug Des 1998; 13:519-528.
- Brown JM. Exploiting the hypoxic cancer cell: mechanisms and therapeutic strategies. Mol Med Today 2000; 6:157-162.
- Connors TA. Bioreductive agents, hypoxic cells and therapy. Eur J Cancer 1996; 32A:1833-1834.
- Phillips RM. Prospects for bioreductive drug development. Exp Opin Invest Drugs 1998; 7:905-928.
- Gutierrez PL. Mechanisms of bioreductive activation. The example of diaziguone (AZQ). Free Radic Biol Med 1989: 6:406-445.
- Denny WA. The design of drugs that target tumor hypoxia. Aust J Chem 2004; 57:821-828.
- deAbreu FC, Ferraz PAdL, Goulart MOF. Some applications of electrochemistry in biomedical chemistry. Emphasis on the correlation of electrochemical and bioactive properties. J Braz Chem Soc 2002; 13:19-35.
- Saijo N. New chemotherapeutic agents for the treatment of non-small cell lung cancer: the Japanese experience. Chest 1998; 113 (Suppl): 17S-23S.
- Lemma K, Sargeson AM, Elding LI. Kinetics and mechanism for reduction of oral anticancer platinum (IV) dicarboxylate compounds by L-ascorbate ions, J Chem Soc. Dalton Trans 2000: 1167-1172.
- Lemma K, Shi T, Elding LI. Kinetics and mechanism for reduction of the anticancer prodrug trans, trans, trans-[PtCl₂(OH)₂(c-C₆H₁₁NH₂)(NH₃)] (JM335) by thiols. Inorg Chem 2000; 39:1728-1734.
- Gandara GR, Lara PN, Goldbert QT, Le QT, Mack P, Lau DHM. Semin Oncol 2002; 29:102-109.
- Hicks KO, Pruijn FB, Sturman JR, Denny WA, Wilson WR. Cancer Res.
- Everett SA, Swann E, Stratford MRL, Naylor MA, Patel KB, Tian N, et al. Modifying rates of reductive elimination of leaving groups from indolequinone prodrugs: a key factor in controlling hypoxia-selective drug delivery. Biochem Pharmacol 2002; 63:1629-1639.
- Wardman P. Free radicals and new prodrugs for cancer therapy. In: Scientific yearbook 2001/2002. London: Cancer Research Campaign; 2001. pp. 36-39.